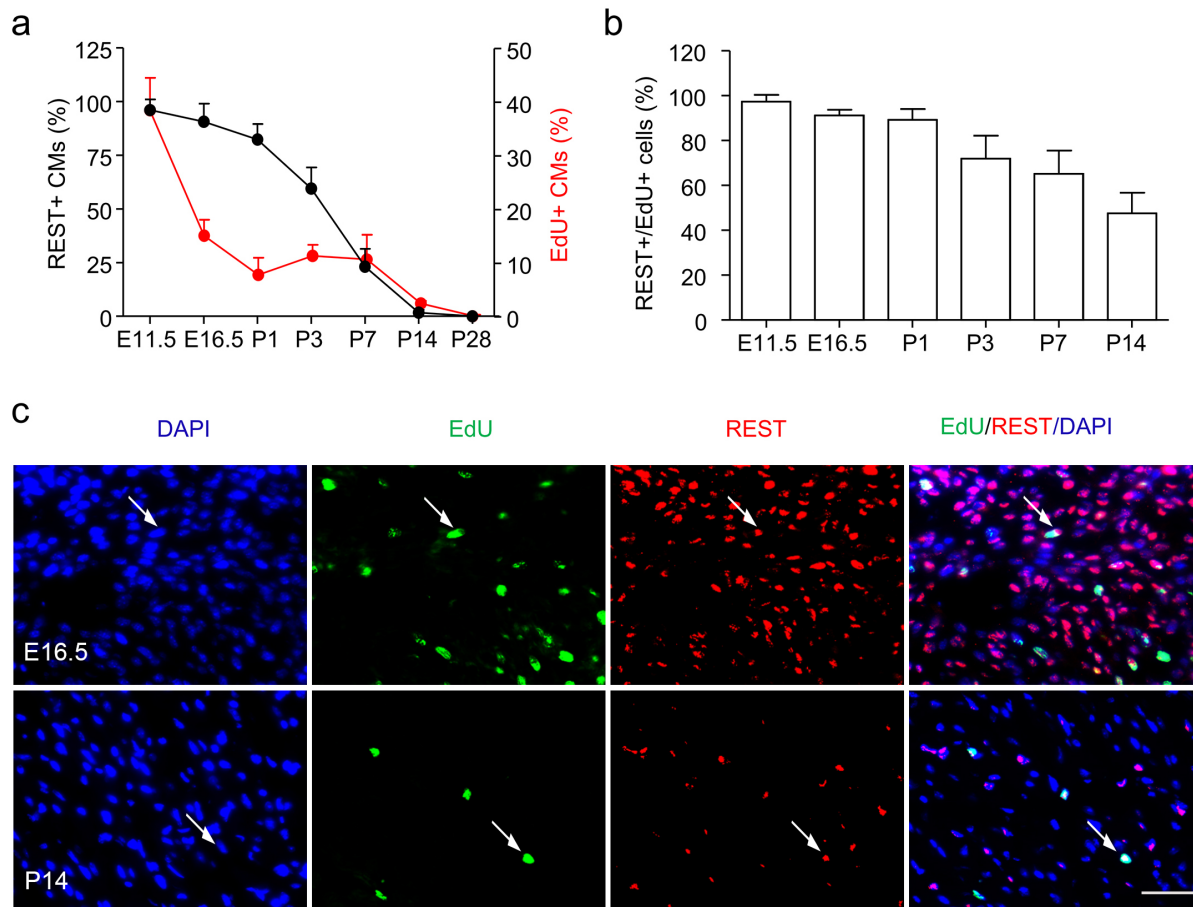
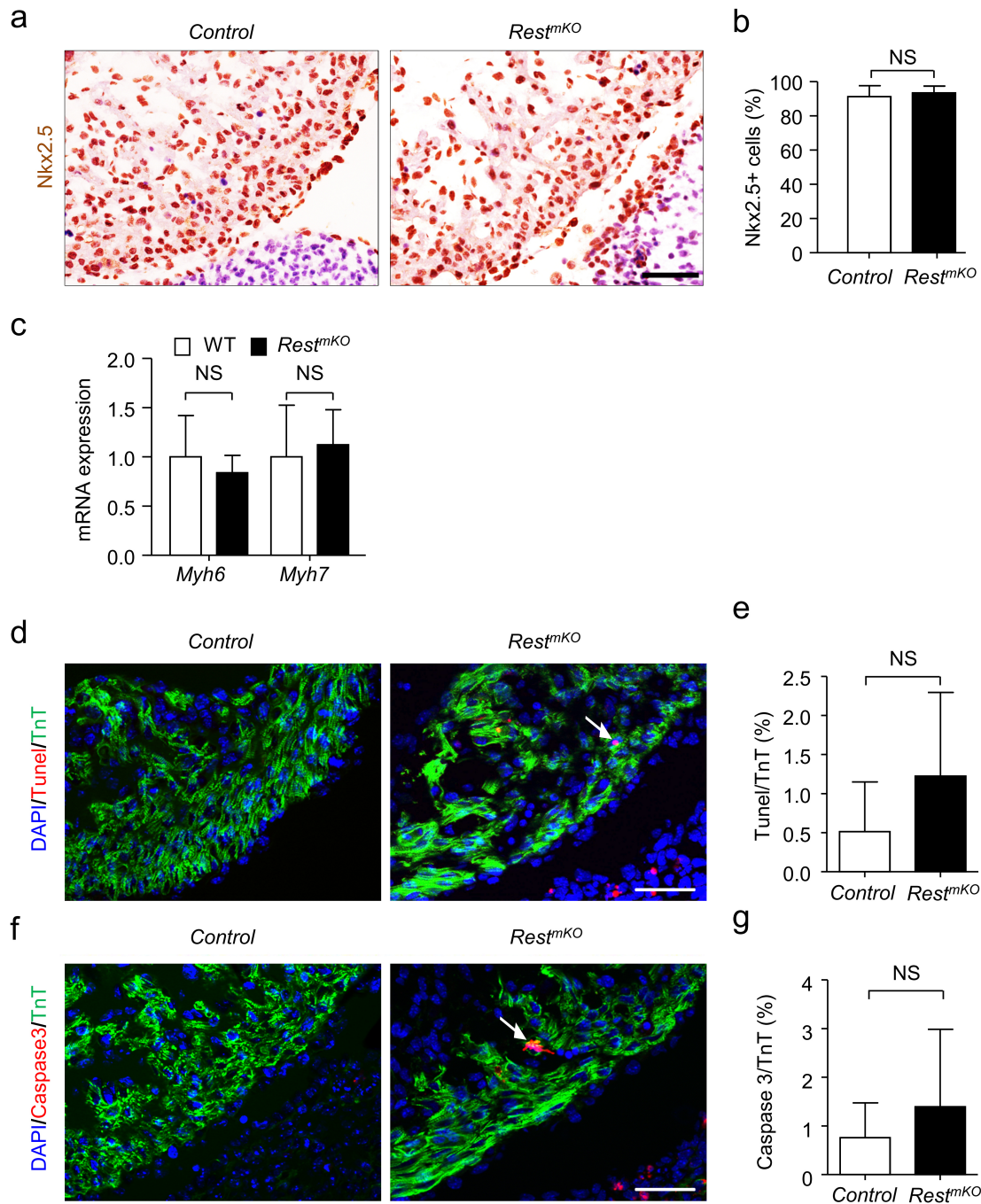


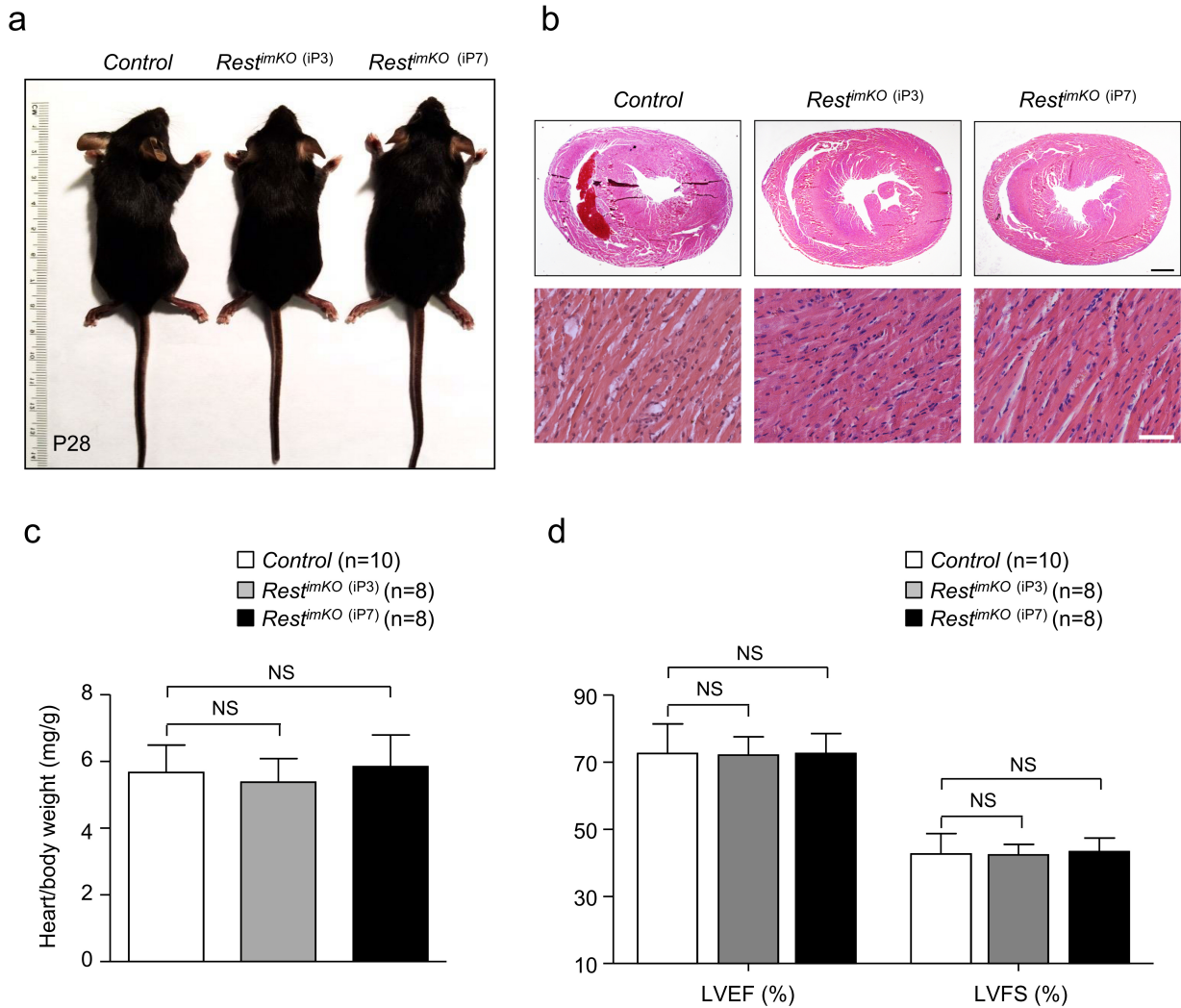
**Supplementary Figure 1. *Rest* expression is developmentally downregulated in embryonic and postnatal hearts.** **a,b**, Western blot analysis showing developmental downregulation of REST. **c,d**, Immunofluorescence showing REST expression in the majority of cardiomyocytes (CMs) of embryonic and neonatal hearts (from E11.5 to P3). After P3 the percentage of REST-expression cardiomyocytes is drastically decreased. Scale bar = 40 $\mu$ m. **e**, qRT-PCR indicating that *Rest* mRNA levels are sustained in the embryonic and postnatal hearts. E11.5 level as one, mean  $\pm$  s.d. (n = 3/stage) in **a,b,e**, n = 6/stage in **c,d**. \* $p$  < 0.05, \*\* $p$  < 0.01 by one-way ANOVA followed by Tukey's test.



**Supplementary Figure 2. REST expression positively correlates with cardiomyocyte proliferation.** **a**, Graph showing developmental changes in the percentage of REST-expressing and EdU+ proliferating cardiomyocytes (CMs). **b,c**, Immunofluorescence revealing a majority of EdU+ proliferating cells expresses REST (arrow). Mean  $\pm$  s.d.,  $n = 6$ /stage for **a**;  $n = 4$ /stage for **b,c**. Scale bar =  $40\mu\text{m}$ .

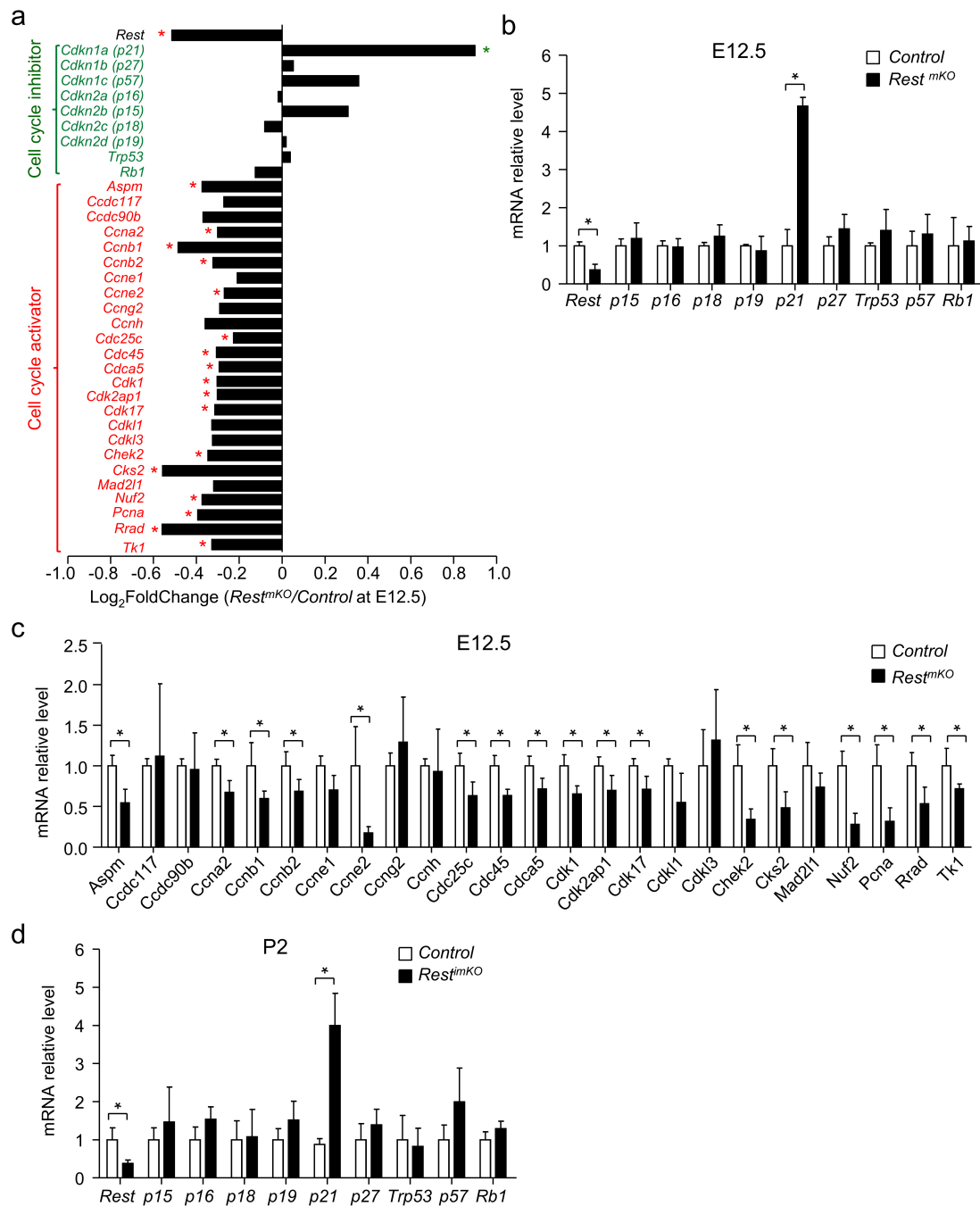


**Supplementary Figure 3. *Rest* is not required for cardiomyocyte differentiation and apoptosis.** **a,b**, Immunostaining showing no difference in the percentage of Nkx2.5-expressing cells in the ventricles between E12.5 control and *Rest*<sup>mKO</sup> hearts. **c**, qRT-PCR analysis indicating no change in the level of *Myh6* and *Myh7* mRNA expression between E12.5 control and *Rest*<sup>mKO</sup> ventricles. **d-g**, Immunofluorescence for TUNEL assay (**d,e**) and Cleaved Caspase 3 (**f,g**) showing no difference in the percentage of apoptotic cardiomyocytes (arrow) in the ventricles between E12.5 control and *Rest*<sup>mKO</sup> hearts. n = 4/group for immunostaining, n = 3 for qRT-PCR, mean ± s.d. NS, no significant by unpaired two-tailed Student's *t*-test. Scale bar = 40µm.

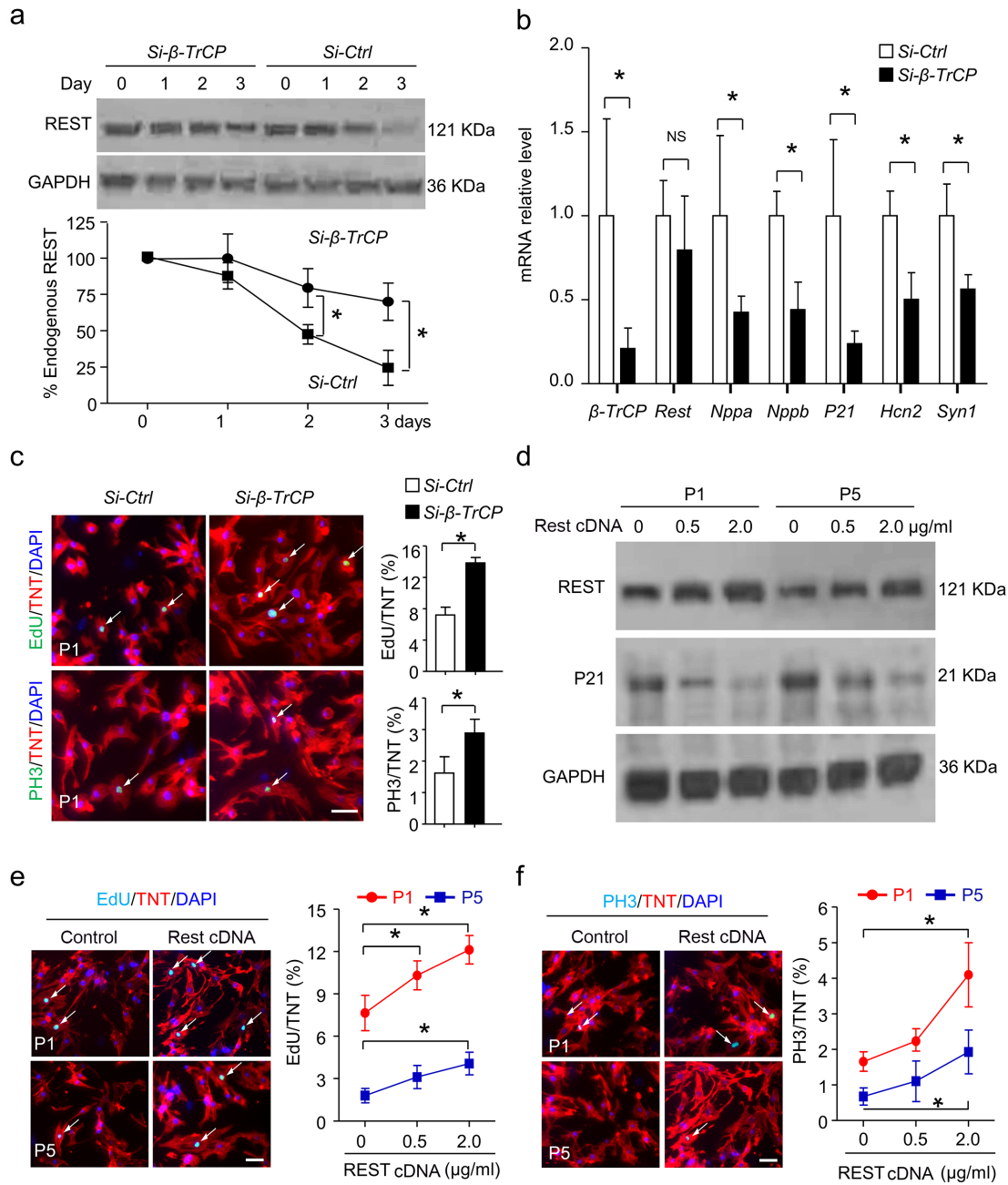


**Supplementary Figure 4. Myocardial *Rest* deletion at P3 or P7 has no effect on cardiac structure and function.** Normal appearance (**a**), cardiac structure by H&E staining (**b**), ratio of heart/body weight (**c**), and cardiac function on echocardiography (**d**) of P28 mice after *Rest* deletion induced at P3 or P7. Mean  $\pm$  s.d. NS, no signification by one-way ANOVA followed by Tukey's test. Controls were Tam-treated *Rest<sup>+/+</sup>;TnT<sup>MerCreMer/+</sup>* mice. Scale bar =100 $\mu$ m or 40 $\mu$ m in the top or bottom panel in **b**.

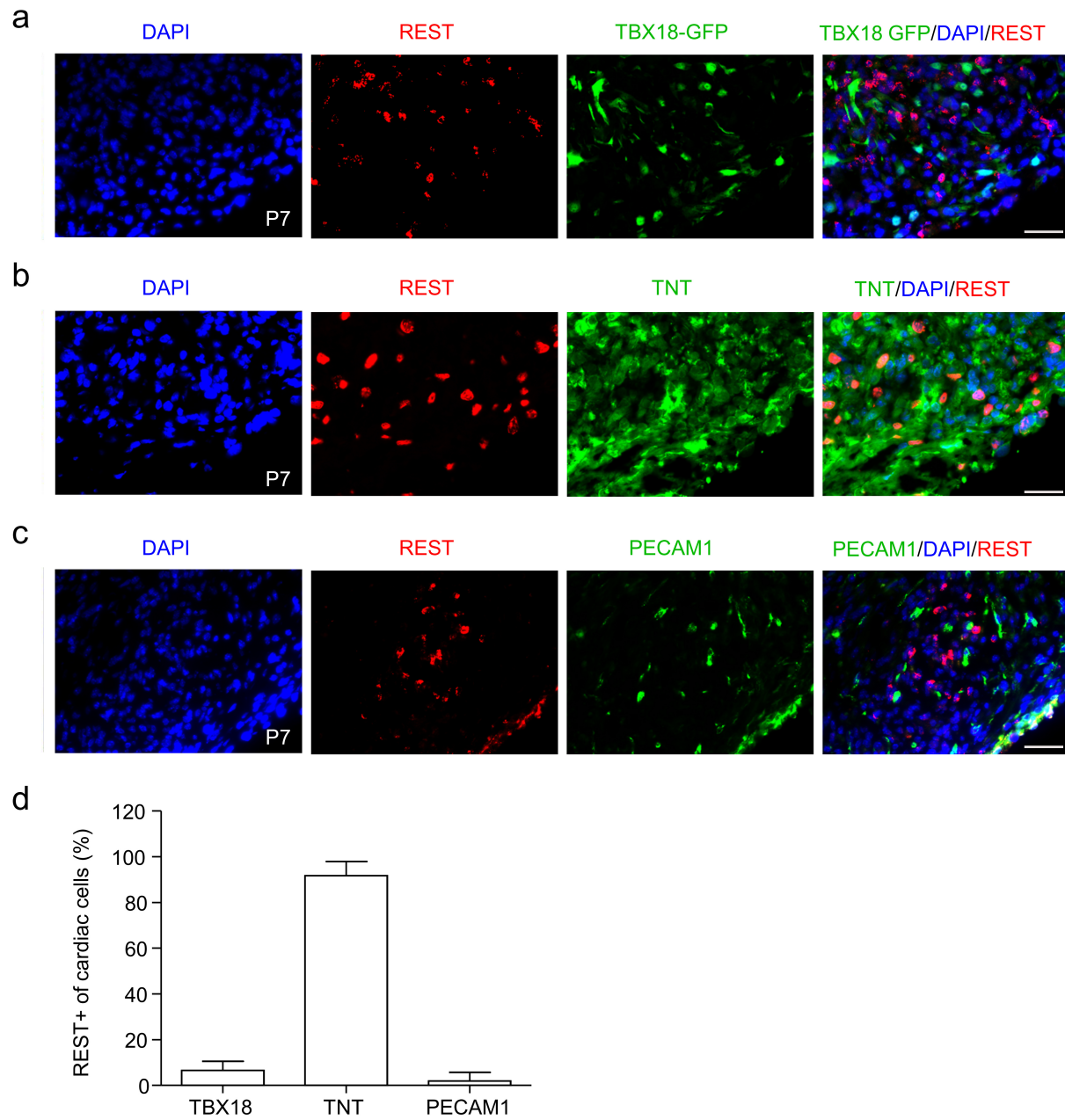




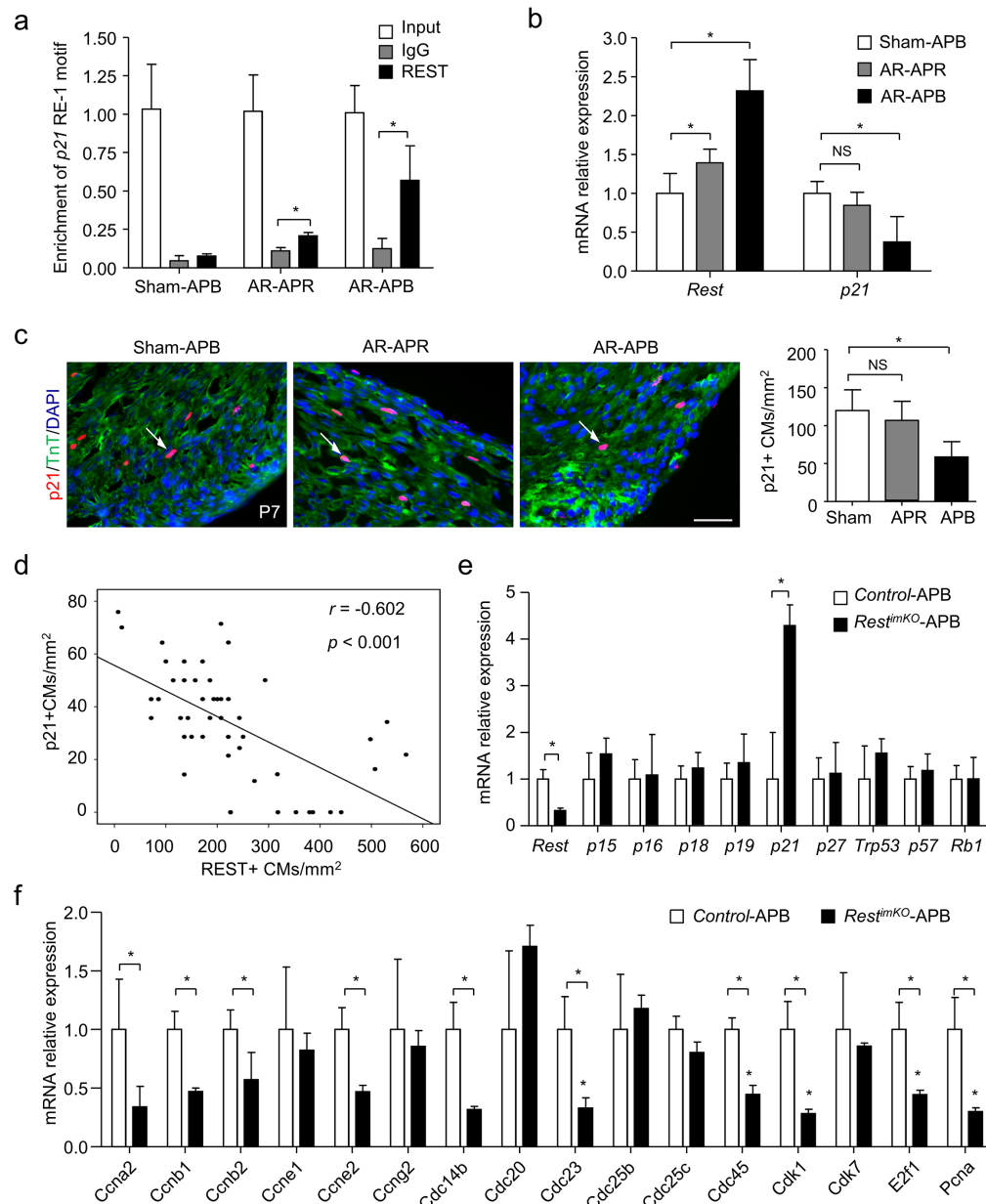
**Supplementary Figure 5. *Rest* deletion results in dysregulated cell cycle gene expression.** **a**, RNA-seq analysis of control versus *Rest* inactivated ventricles of E12.5 hearts showing that among the cell cycle inhibitor genes (green\*), *p21* expression is upregulated, whereas the expression of several cell cycle activator genes is downregulation (red\*). **b,c**, qRT-PCR shows *p21* upregulation (**b**) and downregulation of the cell cycle activator genes (**c**) in E12.5 ventricles with *Rest* inactivation. **d**, qRT-PCR shows *p21* upregulation in P2 ventricles with *Rest* inactivation at P1. Controls were the Tam-treated *Rest*<sup>+/+</sup>; *Tnfr1*<sup>MerCreMer/+</sup> mice. n = 3/group, mean ± s.d. \**p* < 0.05 by unpaired two-tailed Student's *t*-test.



**Supplementary Figure 6. Overexpression of REST induces cardiomyocyte proliferation *in vitro*.** **a-c**, Transfecting siRNA against  $\beta$ -TrCP into the primary cultured cardiomyocytes isolated from P1 hearts stabilizes REST protein (**a**, western blot), downregulates the expression REST target genes (**b**, qRT-PCR), and promotes EdU incorporation and PH3 expression (**c**, immunostaining). **d-f**, Transfecting a plasmid containing Rest cDNA (pHR-NRSF-CITE-GFP, Addgene) into the primary cultured cardiomyocytes isolated from P1 or P5 hearts results in overexpression of REST with reduced p21 expression (**d**, western blot), increased EdU incorporation (**e**) and PH3 expression (**f**). Noted that GFP was not detectable in the assay. Data are presented as mean  $\pm$  s.d. ( $n = 3$ ). \* $p < 0.05$  by unpaired two-tailed Student's  $t$  test in **a-c**; one-way ANOVA followed by Tukey's test in **e, f**. Scale bar = 40  $\mu$ m.

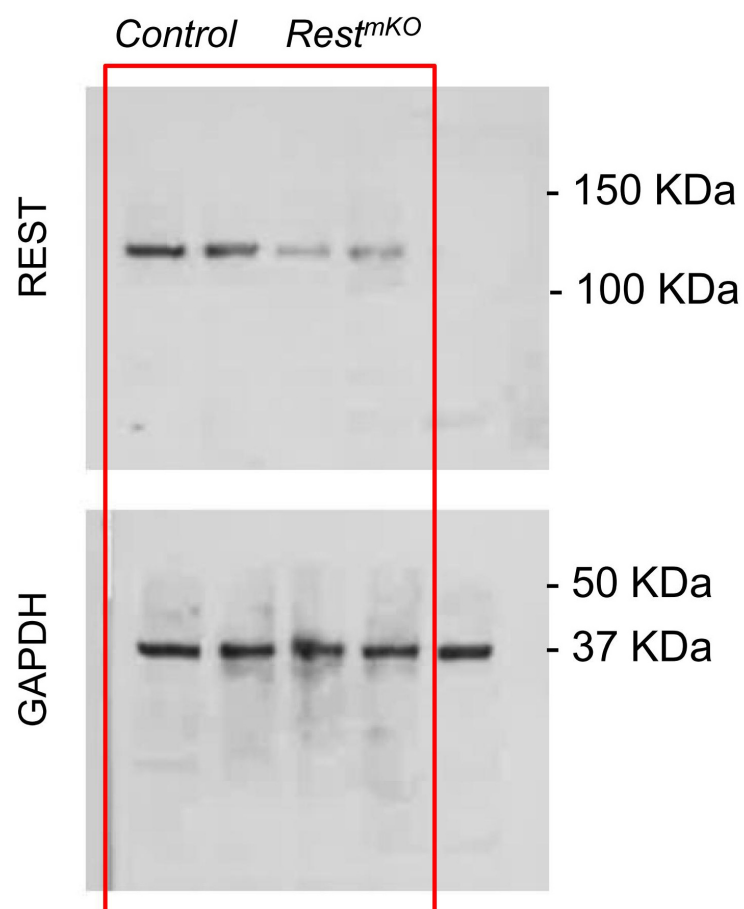


**Supplementary Figure 7. REST re-expresses in cardiomyocytes in regenerating neonatal hearts.**  
**a-d**, Quantitative immunofluorescence showing the majority (~90%) of REST re-expression cells are cardiomyocytes in P7 old regenerating hearts after apex ablation at P4, while a small percentage (<10%) of cells are epicardial or endocardial/endothelial cells. Scale bar = 40 $\mu$ m. n = 4/group, mean  $\pm$  s.d.

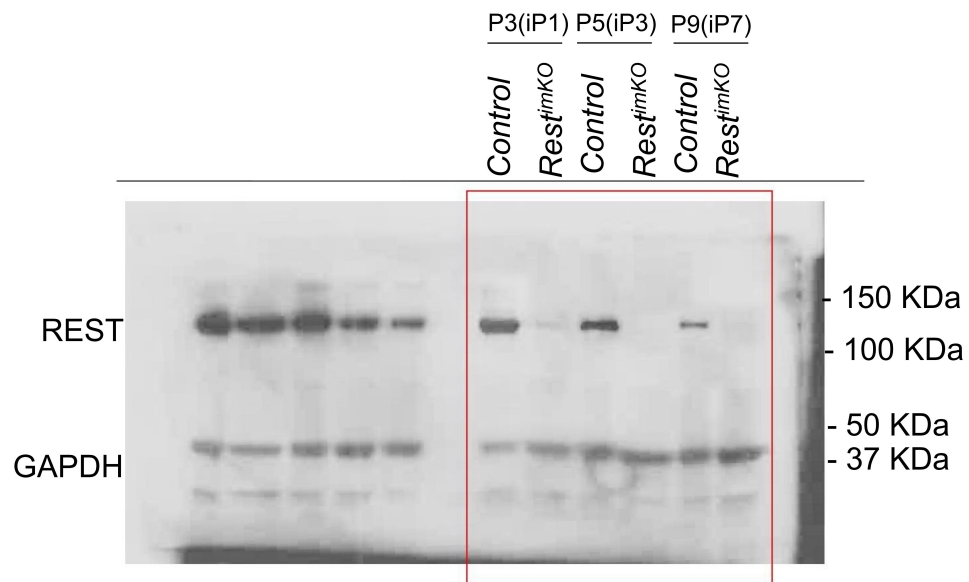


**Supplementary Figure 8. REST represses *p21* expression in regenerating myocardium.** **a**, qChIP indicating increased REST binding to the *p21*-RE1 motif containing DNA fragment in the regenerating apex of P7 hearts. **b**, qRT-PCR showing *Rest* upregulation and *p21* downregulation in the regenerating apex of P7 hearts. **c**, Immunofluorescence revealing decreased number of p21+ cardiomyocytes (CMs) in the regenerative apex of P7 heart,  $n = 4/\text{group}$ . Scale bar =  $40\mu\text{m}$ . **d**, Scatter plots showing a negative relationship of the number of REST+ and p21+ cardiomyocytes. Average counts from 6 fields/section, 3-4 sections/ventricle, 4 ventricles/stage at P7, 14, and 28. **e,f**, qRT-PCR analysis revealing changes in the levels of cell cycle inhibitors (**e**) and activators (**f**) in the regenerating apex resulting from *Rest* deletion.  $n = 3/\text{group}$ , mean  $\pm$  s.d. \* $p < 0.05$  by one-way ANOVA followed by Tukey's test in **a-c**; unpaired two-tailed Student's *t* test in **e,f**. APB/APR, apex border/apex remote.

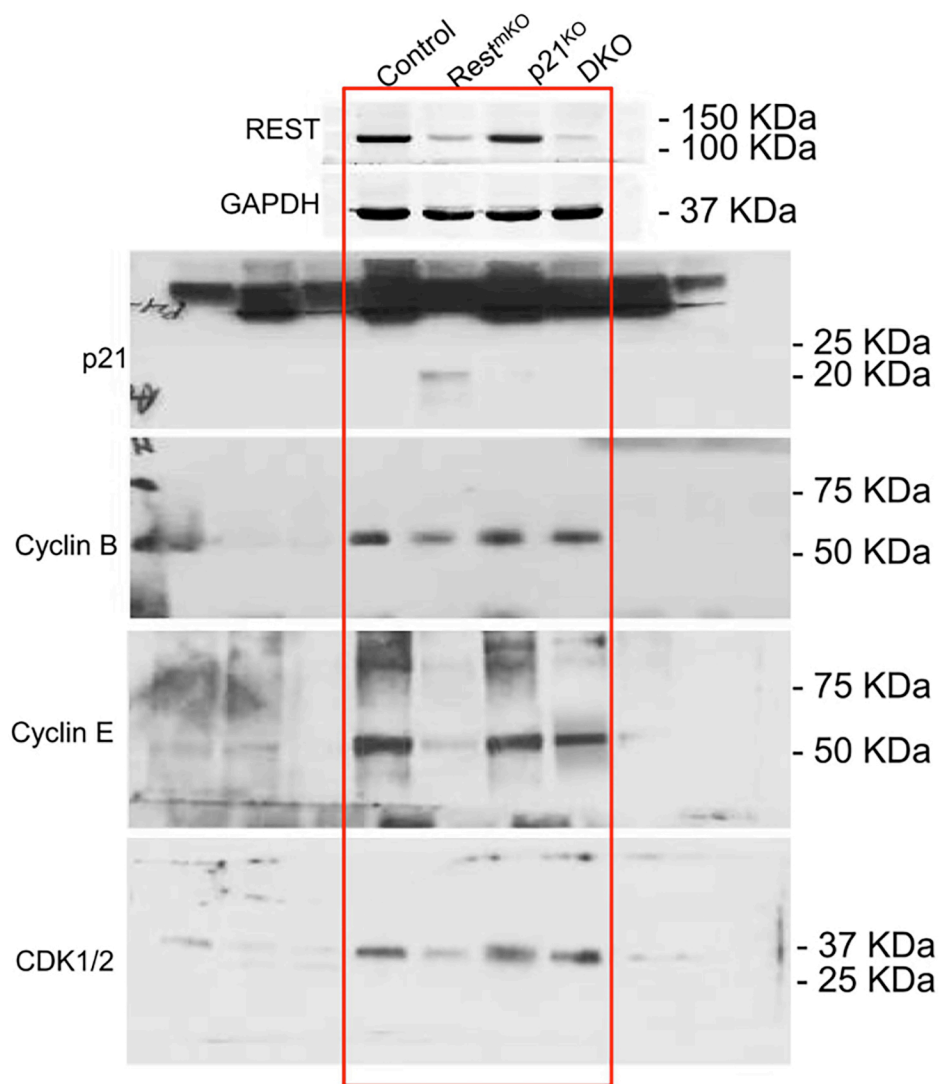




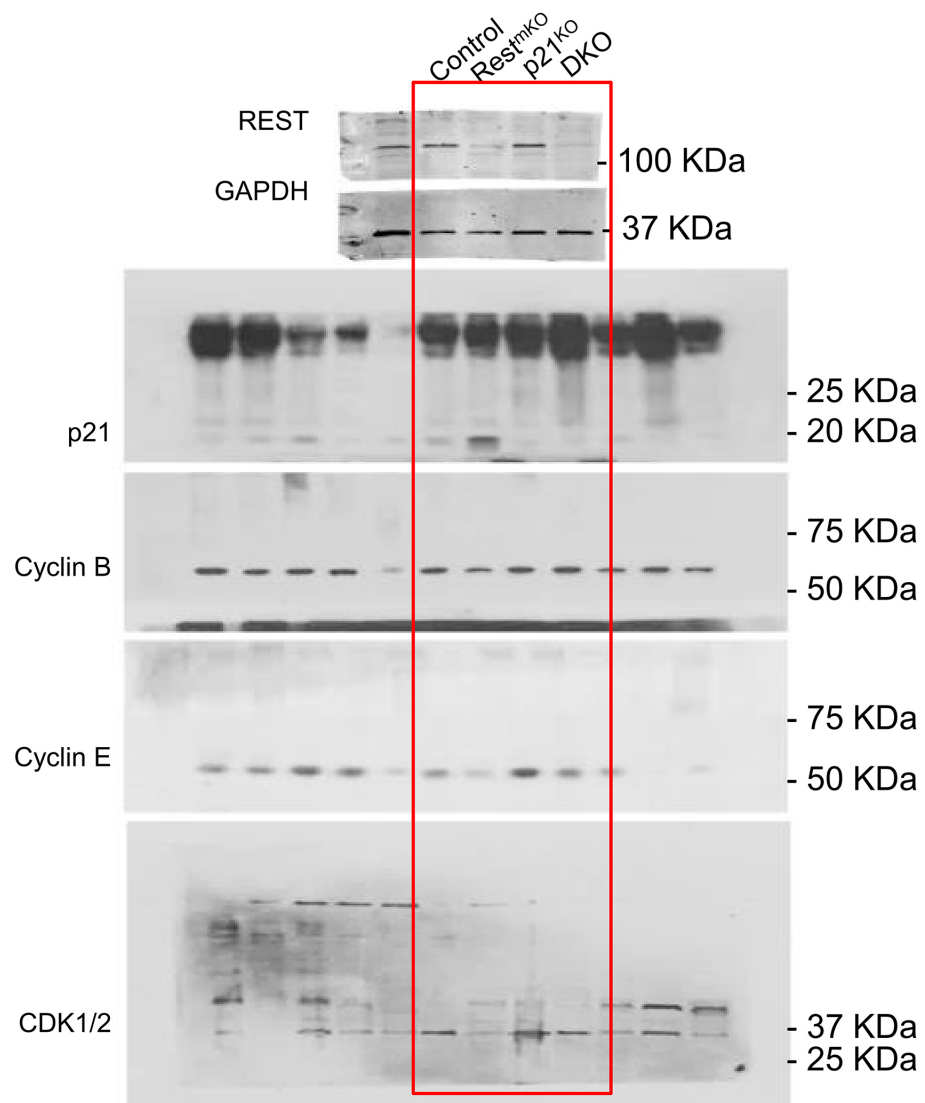
**Supplementary Figure 9.** Full scan of western blots shown in Fig. 1b.



**Supplementary Figure 10.** Full scan of western blots shown in Fig. 2b.

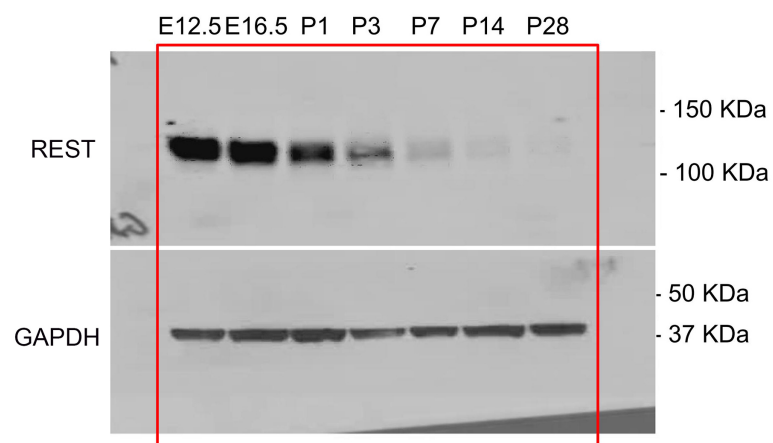


**Supplementary Figure 11.** Full scan of western blots shown in Fig. 4d.

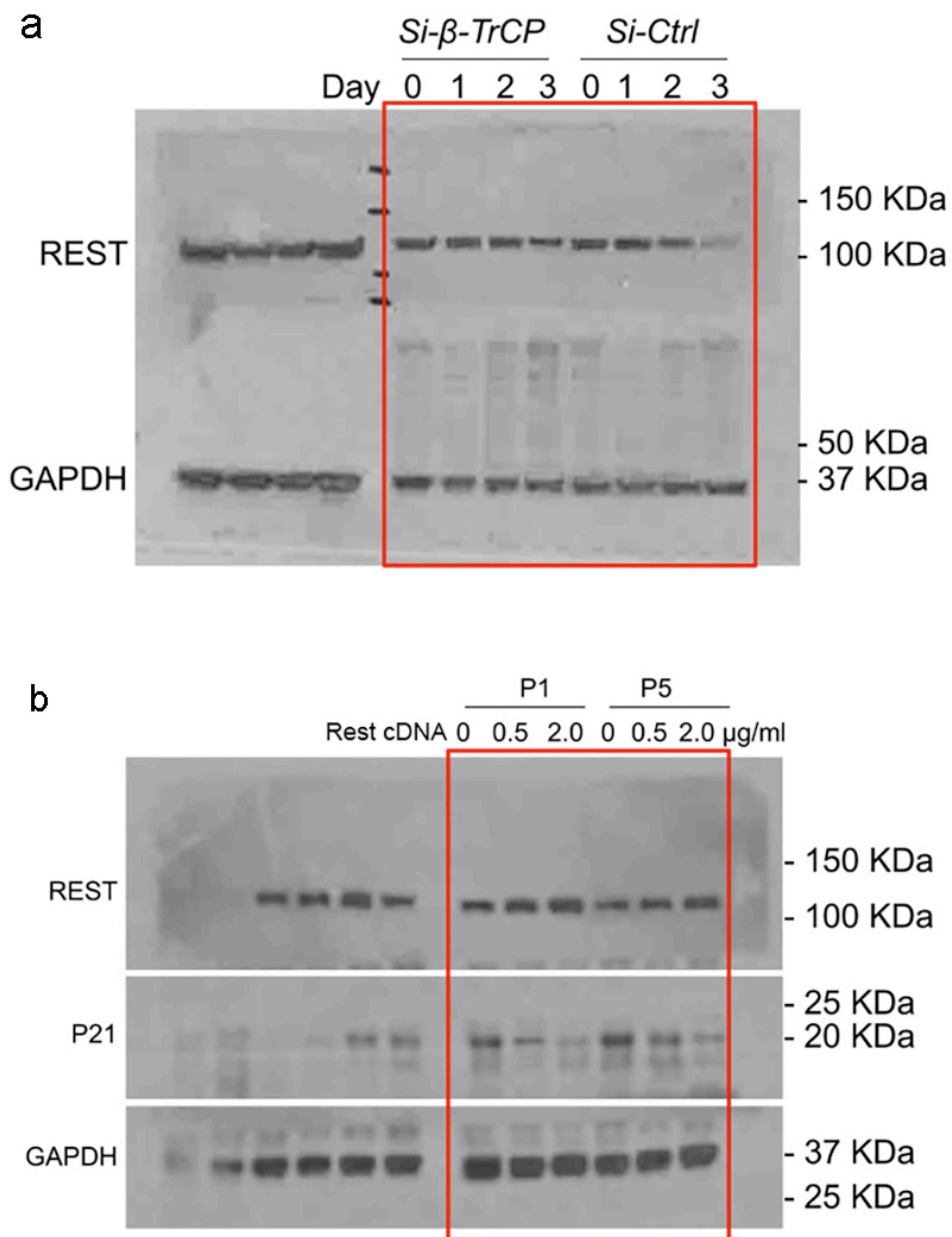


**Supplementary Figure 12.** Full scan of western blots shown in Fig. 5f.





**Supplementary Figure 13.** Full scan of western blots shown in Supplementary Fig. 1a.



**Supplementary Figure 14.** Full scan of western blots shown in Supplementary Fig. 6a (**a**) and 6d (**b**)

**Supplementary Table 1. List of primers used in qRT-PCR, qChIP and luciferase reporter assays**

<b>Assay</b>	<b>Primer sequence (sense 5'-3')</b>	<b>Primer sequence (anti-sense 5'-3')</b>
<b>qRT-PCR</b>		
<i>Aspm</i>	TGGCTATGAGTGAATGCTCTTCC	TCGCGTAAAAACAGTGGCAAG
<i>Ccdc117</i>	GGCTGCAAGAGATTGAGGACA	CCATAGAAGGACGGCTCATAGAC
<i>Ccdc90B</i>	TCCGCTGGACTTCAACTCCT	ACCAACGCATGGGTATCAAAAG
<i>Ccna2</i>	AAGAGAATGTCAACCCCGAAAAA	ACCCGTCGAGTCTTGAGCTT
<i>Ccnb1</i>	CAATTATCGGAAGTGTCGGATCA	CTGGTGAACGACTGAACTCCC
<i>Ccnb2</i>	GCCAAGAGCCATGTGACTATC	CAGAGCTGGTACTTTGGTGTTT
<i>Ccne1</i>	GAAAAGCGAGGATAGCAGTCAG	CCCAATTCAAGACGGGAAGTG
<i>Ccne2</i>	ATGTCAAGACGCAGCCGTTTA	GCTGATTCTCCAGACAGTACA
<i>Ccng2</i>	AGGCTACCCCGGAGAATGATA	ACCTTCATAAGGGCCAAGAATCT
<i>Ccnh</i>	CAGAAACGGCACTGGACCTT	GACCGTGGCATAGCTGGTT
<i>Cdc14b</i>	CTCTACAGCAGACCAAAGAGTG	CCTCAGCATTGTAATGGACTTGA
<i>Cdc20</i>	TTCGTGTTTCGAGAGCGATTTG	ACCTTGGAAGTAGATTTGCCAG
<i>Cdc23</i>	TTCTTCGGTAGTCTCGGTGG	CCAACTCCGCAGACCATT
<i>Cdc25b</i>	TCCGATCCTTACCAGTGAGG	GGGCAGAGCTGGAATGAGG
<i>Cdc25c</i>	GTTCAGCACCCAGTTTTAGGT	AGAATGCTTAGGTTTGCCGAG
<i>Cdc45</i>	TCAGAGGCTGAATGGGTAACA	GGGAAGAACTATGAGGTCTGGT
<i>Cdca5</i>	CGGCGAACACGATCAGGAG	TCCTCGGCCAGATTTAGAGA
<i>Cdk1</i>	TCAAGCACCTGGACACAATTC	GGACCCTTTTGACTTGGTTGG
<i>Cdk17</i>	AACCTTGACGACATCGTTCA	CACGGAGAATTTGGTACAGGAA
<i>Cdk2ap1</i>	GCGACCTCCTCCCAATATCG	GTCTGATCTCTTTCCCAACTCT
<i>Cdk7</i>	GACACCATCCACATTAAAGCC	CACCATACATCCTAGCTCCAAAC
<i>Cdkl1</i>	AGAAGGCTCCTATGGGGTAGT	GGCGATTTTCTTTATGACAGGGT
<i>Cdkl3</i>	TAAGGACACTGGGCGAATAGT	GACCGTGTGGTCAATAAACTCAA
<i>Cdkn1a</i>	GAAAGAAGCGGAAGATCCTCC	GGGCCTCAGGGATTGTTTGG
<i>Cdkn1b</i>	TCAAACGTGAGAGTGCTAACG	CCGGGCCGAAGAGATTCTG
<i>Cdkn1c</i>	GCAGGACGAGAATCAAGAGCA	GCTTGGCGAAGAAGTCGTT
<i>Cdkn2a</i>	CAAAGTGACAGATGCTCCAATCC	TTTTCCTTCTACGGCTCGTTT
<i>Cdkn2b</i>	CCCTGCCACCCTTACCAGA	CAGATACCTCGCAATGTCACG
<i>Cdkn2c</i>	GGGGACCTAGAGCAACTTACT	AAATTGGGATTAGCACCTCTGAG
<i>Cdkn2d</i>	CTGAACCGCTTTGGCAAGAC	GCCCTCTCTTATCGCCAGAT
<i>Chek2</i>	TGACAGTGCTTCCTGTTTACA	AGCTGGACGAACCCTGATACT

<i>Cks2</i>	TCGATGAGCACTACGAGTACC	CCATCCTAGACTCTGTTGGACAC
<i>E2f1</i>	TGCAGAAACGGCGCATCTAT	CCGCTTACCAATCCCCACC
<i>Gapdh</i>	ACGGCAAATTCAACGGCACAGTCA	TGGGGGCATCGGCAGAAGG
<i>Mad2l1</i>	TCCGTCGGAAACCTTTACTAGA	ACTGCCATCTTTCAAGGACTTC
<i>Nppa</i>	ATTGACAGGATTGGAGCCCAGAGT	TGACACACCACAAGGGCTTAGGA
<i>Nppb</i>	AGGGATACAGGAGCTGCTGG	CCTCGCCTCCCAGGCTTCTG
<i>Nuf2</i>	TCCCCAGATACAATGTAGCTGA	CCGGACTCCATACCTAACTGT
<i>Pcna</i>	TTGCACGTATATGCCGAGACC	GGTGAACAGGCTCATTCATCTCT
<i>Rest</i>	GTGCGAACTCACACAGGAGA	GAGGTTTAGGCCCCGTTGTGA
<i>Rb1</i>	GGAACCCAACTCAACGTGAC	TCTGGTCGCTTGTGTGCTC
<i>Rrad</i>	GTCAGAGGAGGGCGTTTACAA	TCCACAGTGATAGAACGGTCA
<i>Syn1</i>	AATATGCGGCTATGGATGTTGG	TCCTGAGTGACCGGAGAGTC
<i>BTRC</i>	AAGACTGTAATAATGGCGAACCC	TCTCTTGTTTATGCAAAGCCTG
<b>qCHIP</b>		
<i>P21</i> -RE1	CACCCCATACTTCCCCTTCT	AAGACCAGGGAATCCCACTT
<b>Luciferase assay</b>		
<i>P21</i> -RE1 WT	CGTGGGTACCCACTTCTTACCTGGGGGTGA	AATGGCTAGCCCATTGCTCAGTGTCTGTG
<i>P21</i> -RE1 Mut1	CTAGGCACTTGCCCCAAAAACAGTGGACTGG	CCAGTCCACTGAGCTGTGGGGTTTTTGCCTAG
<i>P21</i> -RE1 Mut2	CCACAGCTCAGTAAAAAAGAAGGGAATGTATATG	CATATACATTCCCTTCTTTTTTACTGAGCTGTGG
<i>P21</i> -RE1 Del	CTAGGCACTTGCCCCAGAA GGGAATGTATATGC	GCAATACATTCCCTTCTGGGGCAAGTGCCTAG

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